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Thermal changes in young and mature bone nanostructure probed with Ca 2p excitations

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Abstract

Thermal dependence of mineralized bone structure is examined both experimentally by measuring the near edge x-ray absorption fine structure (NEXAFS) and theoretically by applying the 3DSL model to clarify relationship between the local electronic and atomic structure and hierarchical organization of skeleton. The high energy resolution NEXAFS spectra are acquired near the Ca 2p edges in native bone heated from RT up to 450 °C and hydroxyapatite (Ca₁₀(PO₄)₆(OH)₂, OH-Ap) to understand the interplay of short-, long- and super-range order parameters of bone matter. Our focus is on the thermal changes of spectral distribution of oscillator strength for Ca $2p_{1/2,3/2} \rightarrow 3d$ transition in bone and OH-Ap. The investigations have confirmed the assignment of the OH-Ap-to-bone spectral changes to the predicted hierarchy effect on electronic and atomic structure of mineralized bone. At RT the OH-Ap-to-bone red shift δE_{3d} of the transition energy is found equal ≈ 0.2 eV and ≈ 0.1 eV for mature and young bone respectively. We stated that the shift behaves irregular and its magnitude varies from 0.1 eV up to 0.3 eV when the heating temperature grows. Two mechanisms associated with the thermal-induced dehydration of the inter-nanocrystallite spaces and with the subsequent atomic restructuring of the nanocrystallite interface in mineralized phase are revealed. We have detected that the OH-Ap-to-bone red shifts of the Ca $2p \rightarrow 3d$ transition in young bone are smaller than those of in mature bone. The origin of the age differences is discussed.

1. Introduction

Hierarchical structuring provides fascinating mechanical properties and high material functionality of bone tissue [1–9]. Studying it one can achieve deeper understanding of the hierarchical and self-assembled materials. To date, many efforts have been invested to investigate the role of chemical composition in mechanical and physical properties of bone [10–12]. These properties are also tightly related with the skeletal hierarchical organization. Regrettably the hierarchy effect on bone tissue in nanoscale remains without sufficient attention. The hard hydroxyapatite (OH-Ap, $Ca_{10}(PO_4)_6(OH)_2$) strata in combination with the flexible collagen fibrils make bone more elastic and firm [3–6] than the initial mineral and the collagen molecule. Water is the third basic component [1, 4, 6], the role of which in physical, chemical and mechanical properties of bone tissue is not completely understood [1, 5, 13–15].

Medical treatment and rehabilitation of patients with skeletal pathology is of great socio-economic importance. In particular, osteoarthritis is the fourth most common cause of hospitalization [16]. Material science, biological and medical investigations trace the complicated hierarchy of the skeleton designs from macro- to nanolevel [1–9, 17, 18]. Nanolevel research encounters especially great difficulty because local electronic and atomic structure of bone is not sufficiently studied. This knowledge gap prevents solutions of many fundamental and applied problems, such as bio-design of advanced materials [3–9, 16, 19, 20], controlled ion exchange and paleontological research [21, 22]. Nanolevel studies open a perspective way for medical imaging and nanodiagnostics of bone tissue in normal and pathological conditions.

Recently, theoretical and x-ray spectroscopic investigations are carried out to understand relationship between hierarchical organization of native bone and its local electronic and atomic structure [18]. According to the 3D superlattice (3DSL) model [18] this structure is a subject of complicated interplay of the short-, long- and super-range order parameters of bone material. In particular, the interplay results in the OH-Ap-to-bone red shift of the density of electronic states and the core–shell to valence shell transitions. The predicted OH-Ap-to-bone spectral changes are proportional to the ratio of the hydrated layer thickness to the linear size of the nanocrystallites of OH-Ap (NOH-Ap) and specify the hierarchy effect on electronic and atomic structure of bone.

Comparison of the experimental near-edge x-ray absorption structure (NEXAFS) spectra of native bone at the Ca 2p, P 2p and O 1s thresholds with the known spectra of OH-Ap has confirmed the prediction. The red shifts of $2p \rightarrow 3d$ absorption transitions extracted from the experimental Ca L_{2,3} NEXAFS analysis are found varying from 1 eV up to 0.2 eV [18, 23]. In general these values agree with the model shift $\approx 0.4 \text{ eV}$ computed in assumption that empty spaces separate the equal NOH-Ap in their coplanar assembly. Note that this estimate gives the maximal value of the red shift [18]. To understand why the larger shifts have been detected the authors of [24] have pointed out that the extracted shifts are obtained from the NEXAFS spectra of bone samples by using different techniques to prepare them and different methods to calibrate the absolute energy of x-ray transitions.

In the present work to avoid these uncertainties our main goal is to measure more accurately the Ca 2p1/2,3/2 NEXAFS spectra of bone and OH-Ap under the same experimental conditions. Besides, we focus on thermal-induced changes in bone tissue to study in more detail the hierarchy effect on electronic and atomic structure of mineralized bone and to check the applicability of the 3DSL model for its description. To probe the thermal properties of native bone the high resolution Ca 2p1/2,3/2 NEXAFS spectra of bone samples heated from RT up to 450 °C are measured and examined. Since the thermodynamic equilibrium in them is not realized as the collagen and water networks are subjects of substantial structural changes the heating temperature T and heating time τ are used to specify the thermal loads. In the work the time period τ is fixed whereas the temperature varies. Thus, the T dependence of the Ca 2p_{1/2,3/2} NEXAFS is used to probe the thermal properties of native bone.

Heating up to 200 °C leads to removing of the inter-crystallite water from mineralized phase [11, 25]. The microthermal analysis and atomic force microscopy of collagen matrix [26] evidences that 65 °C, 130 °C–180 °C, 260 °C, 340 °C and 420 °C are the transition temperatures for denaturation of native

hydrated collagen, evaporation of bound water and denaturation of calcified collagen, conformation changes of the collagen molecules from triple helix structure to random coil, as well as the first and second stages of collagen matrix degradation, respectively. Taking into account the essential structural changes in the collagen fiber and water networks and that thermal loads play a crucial role in hierarchical nanostructures [27] a complicated T-dependence of Ca 2p NEXAFS spectra of mineralized bone is expected. The 3DSL model also predicts irregular thermal changes in electronic and atomic structure of mineralized bone tightly related with the super periodicity of the coplanar assemblies of nanocrystallites.

The experimental details of our spectroscopic studies are described in the section 2. The T-dependences of the Ca 2p NEXAFS spectra and the OH-Ap-to-bone red shifts of the 'white' lines are measured for young and mature bone tissue and presented in section 3. To rationalize the observed thermal-induced and agerelated changes in mineralized bone the 3DSL model is applied. The dehydration and restructuring mechanisms resulting in the spectral shift and line shape distortion of the Ca 2p excitations in bone matter are discussed in section 4.

2. Experimental

All absorption measurements were carried out using monochromatic synchrotron radiation at the RGBL beamline of the BESSY II synchrotron light source of Helmholtz-Zentrum Berlin. Powdered bone and synthetic apatite samples were used as objects. The bone samples are prepared from cortical middle third of the femur, tibia and humerus white mongrel mature male rats weighting 250–280 g and young (prenatal) rats weighting 40–60 g. Cortex has been (i) thoroughly cleaned of soft tissue, (ii) washed in saline, (iii) dried with blotting paper, and (iv) grinded to fine powder with particles of size ~1 μ m.

The hydroxyapatite of almost stoichiometric composition (a = 9.416 (2) Å, c = 6.880 (1) Å) was synthesized by reverse precipitation method in an ammonium-containing solution. An aqueous solution containing ammonium hydrogen phosphate $((NH_4)_2HPO_4)$ (100 ml 0.1 mol l⁻¹ (10 mmol of phosphate ions)) was dropped by drop into a stirred solution of calcium nitrate (Ca(NO₃)₂) (200 ml 0.1 mol l^{-1} (20 mmol of calcium ions) heated to 85 °C-90 °C. Stirring was carried out continuously for 30 min. The required pH value = 8-11 was maintained by adding concentrated ammonium hydroxide (NH₄OH) solution to the ammonium hydrogen phosphate $((NH_4)_2HPO_4)$ solution. The resulting solution with a fine-crystalline white precipitate was kept at $T = 90 \degree C-100 \degree C$ for 2 h. The precipitate was washed by distilled water several times, dried at 100 °C-110 °C for 24 h, and then grinded in a mortar until the sample

was homogenized. The ratio of the calcium cations to the phosphate ions in the solution was at least 2/1throughout the entire synthesis process to ensure the crystallization of apatite with the Ca/P ratio ~1.67. X-ray diffraction measurements were used to control the atomic structure of synthetic samples. This synthesis method is described in more detail in [28].

The samples for x-ray absorption measurements were prepared ex situ in air: the bone and gold powders were mixed and pressed to prepare the pellets. Gold powder was added to reduce the charging of samples during their irradiation with a very intense synchrotron radiation beam. In addition, the samples of the bone tissue and a reference compound (synthesized OH-Ap nanopowder) were prepared by rubbing the bone powders into a scratched surface of a clean substrate (metallic gold sheet $5 \times 5 \text{ mm}^2$ in size). The samples were fixed on the holder and introduced into the preparation chamber, where they can be heated in the furnace for 30 min at various fixed temperatures T in the range from RT to 450 °C. The sample holder temperature was measured using a S-type thermocouple. After the heating the samples were transferred into the measuring chamber.

The samples were placed at the angle of ~45° with respect to the incident beam of the monochromatic radiation and the size of the focal spot on the sample was around $1 \times 1 \text{ mm}^2$. The NEXAFS spectra were measured by recording the total electron yield (TEY) as a function of incident photon energy $h\nu$. The TEY proportional to the absorption cross section [29, 30] was acquired by measuring the drain current from the sample and characterized by probing depths of >10 nm. X-ray absorption investigations were performed in the measuring chamber under ultrahigh vacuum conditions with a residual gas pressure of ~10⁻⁹ mbar. No remarkable charging or decomposition effects for all the samples due to their irradiation with the intense beam of soft x-rays were detected.

The NEXAFS spectra were recorded with a photon-energy resolution of 78 meV for the Ca 2p edge ($h\nu \sim 350$ eV). The photon energy $h\nu$ over this range was calibrated by measuring Au 4 f_{7/2} photoelectron peak using photons of the 1st and 2nd orders of diffraction with energies of 355 eV and 710 eV, respectively. The energy positions of the Ca 2p absorption structures of the bone samples were determined with an accuracy of about 0.1 eV.

3. Thermal dependence

The Ca 2p NEXAFS spectra of the mature and young bone measured at the different heating temperature T are plotted in figures 1 and 2. Close similarity of the x-ray absorption spectra of bone and OH-Ap is evident. In all spectra the intense resonances A and B and the weak structures located on their low-energy sides specify photoabsorption in the spectral interval

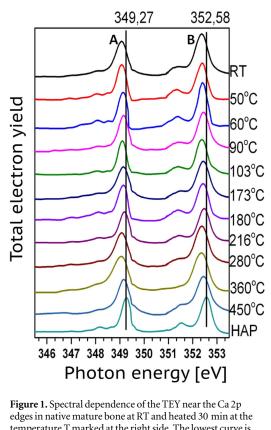


Figure 1. Spectral dependence of the 111 hear the Ca 2p edges in native mature bone at RT and heated 30 min at the temperature T marked at the right side. The lowest curve is the reference Ca 2p NEXAFS spectrum of OH-Ap. To visualize the OH-Ap-to-bone red shifts the energy positions of the 'white' lines A and B in OH-Ap are shown by the vertical lines.

 $346 < h\nu < 354$ eV. The NEXAFS spectra demonstrate a typical spectral distribution of oscillator strength for x-ray transition near the Ca $2p_{1/2,3/2}$ edges in various calcium compounds such as $Ca_3(PO_4)_2$, CaCO₃ [23, 31–35]. The spectral distribution is dominated by multiplet splitting of the Ca²⁺ 2p⁵3d¹ electronic configuration of the $2p \rightarrow 3d$ transition final state in the crystal field. The theoretical investigations [31, 32] have assigned the absorption lines A and B with the splitting $Ca^{2+} 2p^5 3d^1(a_2)$ and $2p^5 3d^1(b_2)$ components. Their intensities and energy positions depend strongly on the surrounding potential and its point group symmetry. Since they are subjects of changes in bone tissue the labels a_2 and b_2 are not regarded here as the corresponding irreducible representations.

By examining the experimental spectra in figures 1 and 2 the distinct deviations of the Ca 2p excitations of bone in comparison with OH-Ap can be revealed. First of all we observe the downward energy shift of the absorption lines A and B relative to their reference positions in the Ca 2p absorption spectra of OH-Ap. In the figures the vertical solid lines trace the measured positions 349.3 eV and 352.6 eV of the Ca 2p excitations in OH-Ap. The OH-Ap-to-bone red shifts $\delta E_{A(B)}$ of the A and B lines are defined as

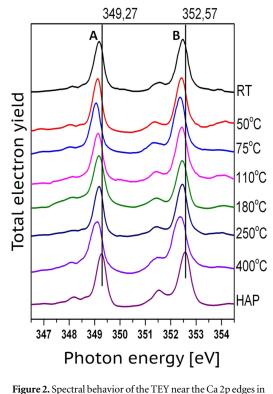


Figure 2. Spectral behavior of the TEY near the Ca 2p edges in young bone heated 30 min at the temperature marked from the right side. The lowest curve is the reference spectrum of OH-Ap. The energy positions of the white lines A and B in OH-Ap are shown by the vertical lines to visualize the OH-Ap-to-bone red shifts.

Table 1. The OH-Ap-to-bone red shifts and changes in FWHM of the Ca 2p excitations A and B of mature bone as a function of heating temperature T.

| T(°C) | δE_A^m (eV) | $\delta E_B^m ({ m eV})$ | (eV) | (eV) |
|-------|---------------------|--------------------------|------|------|
| RT | 0.2 | 0.2 | +0.2 | +0.2 |
| 50 | 0.2 | 0.2 | 0 | 0 |
| 60 | 0.2 | 0.2 | -0.1 | -0.1 |
| 90 | 0.1 | 0.1 | +0.1 | +0.2 |
| 103 | 0.2 | 0.3 | 0 | +0.1 |
| 173 | 0.2 | 0.2 | +0.3 | +0.2 |
| 180 | 0.1 | 0.2 | 0 | 0 |
| 216 | 0.1 | 0.1 | +0.1 | +0.1 |
| 280 | 0.3 | 0.3 | +0.2 | +0.2 |
| 360 | 0.3 | 0.3 | +0.4 | +0.2 |
| 450 | 0.1 | 0.1 | +0.2 | +0.4 |
| OH-AP | 0 | 0 | 0 | 0 |

$$\delta E_{A(B)} \equiv E_{OH-Ap}^{(A(B))} - E_{bone}^{(A(B))},\tag{1}$$

where $E_{OH-Ap}^{(A(B))}$ and $E_{bone}^{(A(B))}$ are the energy positions of the lines A or B in OH-Ap and native bone respectively. Both the sign and the mean value of the shift $\delta E_{A(B)}$ agree reasonably with our expectations based on the 3DSL model and support the hierarchy effect on the core-to-valence transition energies in mineralized bone predicted and discussed in [18, 24].

By comparing the Ca 2p NEXAFS spectra we also see the distinct thermal-induced and age-related deviations in them. The measured OH-Ap-to-maturebone δE_A^m and δE_B^m and OH-Ap-to-young-bone δE_A^y

Table 2. The OH-Ap-to-bone red shifts and changes in FWHM of the Ca 2p excitations A and B of young bone as a function of heating temperature T.

| T (°C) | $\delta E_A^{\gamma}(\mathrm{eV})$ | δE_B^{γ} (eV) | $\delta W_A^{\gamma}(\mathrm{eV})$ | $\delta W_B^{\gamma}(\mathrm{eV})$ |
|--------|------------------------------------|----------------------------|------------------------------------|------------------------------------|
| Т | 0.1 | 0.1 | +0.1 | +0.2 |
| 50 | 0.2 | 0.2 | 0 | 0 |
| 75 | 0.2 | 0.2 | +0.1 | +0.2 |
| 110 | 0.1 | 0.1 | +0.1 | +0.1 |
| 180 | 0.1 | 0.1 | +0.2 | 0 |
| 250 | 0.1 | 0.1 | +0.1 | +0.1 |
| 400 | 0.2 | 0.2 | +0.2 | +0.2 |
| OH-AP | 0 | 0 | 0 | 0 |

and δE_B^{γ} red shifts of the Ca 2p excitations as a function of T are collected in tables 1 and 2. The shifts δE_A^m and δE_B^m vary from 0.3 eV up to 0.1 eV; the full-width-athalf-maximum (FWHM) demonstrates narrowing up to -0.1 eV and broadening up to +0.4 eV. Considering the spectra of young bone we see that the thermalinduced changes of both the red shifts and FWHM do not exceed 0.2 eV.

The OH-Ap-to-bone red shifts for young (*y*) and mature (*m*) bone averaged over the thermal interval are found $\delta E_A^m \approx 0.18 \text{ eV}$, $\delta E_B^m \approx 0.2 \text{ eV}$, $\delta E_A^y \approx \delta E_B^y \approx 0.14 \text{ eV}$. So, the shifts of the Ca $2p \rightarrow 3d$ transitions in young bone are smaller than those of in mature bone. Note, that the detected shifts $0 < \delta E_{A(B)} \leq 0.3 \text{ eV}$ are in good agreement with the 3DSL model and with the experimental results presented in [18, 24].

Analyzing the thermal dependence of the core-tovalence transitions in mature bone we see that the shift δE_A^m comes through maximums $\approx 0.3 \text{ eV}$ at 280 °C– 360 °C and the shift δE_B^m reaches the maximums $\approx 0.3 \text{ eV}$ at lower T ≈ 103 °C and in the interval 280 °C–360 °C too. The both shifts come through the minimum $\approx 0.1 \text{ eV}$ at T ≈ 90 °C, 216 °C and 450 °C. The OH-Ap-to-mature-bone changes in the FWHM demonstrate also the distinct T-dependence. These 'white' lines become narrower when T increases from RT up to 60 °C and broader at higher temperatures. The maximal broadening $\delta W_A^m \approx \delta W_B^m \approx 0.4 \text{ eV}$ is observed at 360 °C and 450 °C.

As for the Ca 2p excitations of young bone the maximal shifts $\delta E_A^{\gamma} \approx \delta E_B^{\gamma} \approx 0.2$ eV are reached twice at 50 °C–75 °C and 400 °C. Their maximal narrowing and broadening is observed at 50 °C and 400 °C respectively.

Since the $Ca^{2+}2p$ hole relaxation in native bone and OH-Ap is nearly the same the Ca 2p NEXAFS spectra evidence that the density of $Ca^{2+}3d$ states in bone is distinctively shifted downward relative to the 3d-band in OH-Ap and demonstrates the specific thermal and age changes. These changes cannot be evidently rationalized in terms of the thermal phonon distribution in OH-Ap (see, e g. [36]). On this background we infer that the observed thermal-induced and age-related OH-Ap-to-bone variations of the Ca 2p excitations are motivated by the composite nature of bone and tightly related with its hierarchical organization. By analyzing the changes we bring forward a suggestion that they are primarily controlled by dehydration and thermal-induced chemical reactions occurring in the hydrated-layers and nanocrystallite interfaces.

The thermal shifts $\delta E_A^{m(y)}(T)$ and $\delta E_B^{m(y)}(T)$ as well as the changes $\delta W_A^{m(y)}(T)$ and $\delta W_B^{m(y)}(T)$ are similar in magnitude. But they are not identical functions. In the low temperature regime (from RT up to $\approx 50^\circ$ -60 °C) one may see the narrowing of the A and B lines when T grows. In some cases the FWHM is narrower than that of in OH-Ap. This extra narrowing can be understood within the 3DSL model by combining the supertlattice and dehydration effects on electronic and atomic structure of bone [18]. For higher temperatures (T > 250 °C) the 'white' lines become broader. We assign this broadening to the atomic restructuring as the number of non-equivalent calcium positions increases.

4. Discussion

Transmission electron microscopy and small angle x-ray scattering on bone makes it evident that intraand inter-fibrilar associations of OH-Ap-nanocrystallites form a series of parallel and spiral wound mineralized plates [5, 10, 18, 37]. These plates are usually described as coplanar assemblies of NOH-Ap submerged into the saturated aqueous solution containing Ca²⁺, Mg²⁺, OH⁻, $[PO_4]^{3-}$, $[CO_3]^{2-}$, and other ions [7, 10, 18, 37]. Both the size distribution of nanocrystallites and their elemental composition in the plates differ in normal and pathological conditions [10, 11, 17, 22, 38–40]. The interplay of the 3D atomic structure of OH-Ap with the higher lying levels of the hierarchical organization of bone determines its nanolevel morphology.

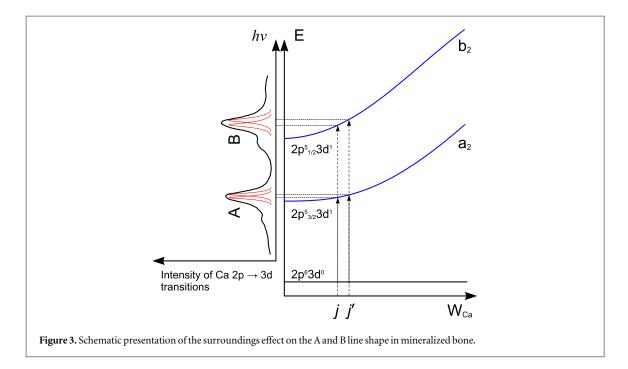
The electronic and atomic structure of an individual mineralized plate is described as a 3D-superlattice assembled from the uniformly sized crystallites surrounded by the hydrated nanolayer [18]. The size of the effective NOH-Ap and the number of atoms per the crystallite can be approximated as $20 \times 7.5 \times 3.5 \text{ nm}^3$ and 40 000 respectively [17]. The advantage of the 3DSL model is the ability to link the band structure of the single-crystal OH-Ap (see, e.g. [41, 42]) with the electronic structure of native bone taking its hierarchical organization into account. This approach promotes a deeper understanding of the electronic and atomic properties of bone matter.

Considering the electronic and atomic structure of the single-crystal OH-Ap the main regularities inherent to the physical and chemical properties of the hierarchical nanostructure can be predicted and extracted. In particular, the electronic structure of the mineralized bone and its thermal properties can be described and computed by solving the Heine equation [18, 43]. Its solutions are determined by thermal dependence of the coefficient Re(1/TT') where T and T' are the amplitudes of electron transmission through the NOH-Ap and the hydrated nanolayer respectively. The amplitude T' attracts our special attention as it is regarded to be tightly related with the thermal-induced dehydration and the subsequent restructuring of the NOH-Ap interface. Thus, by examining the T-dependence of T' we have a chance to disentangle the thermal properties of mineralized phase and rationalize the observed OH-Ap-to-bone changes $\delta E_{A(B)}^{m(y)}$ and $\delta W_{A(B)}^{m(y)}$ as functions of thermal load.

The Ca and P 2p and O 1s NEXAFS analysis performed in the work [18] has shown that at RT the ions Mg^{2+} , Ca^{2+} , $(OH)^{-}$ and $(PO)_4^{3-}$ surrounded by the solvation shells (H₂O)_n play an important role in forming the electronic properties of mineralized bone. These shells shield effectively the columbic potential of the ions and weaken their chemical bonding with other atoms in the inter-crystallite space. The thermal treatment as it is stated in [11, 25, 26, 44] leads to the denaturation of mineralized collagen and to the removal of inter-crystallite water and, hence, to the unfolding of the solvation shells. This makes the columbic shielding less effective and induces restructuring of the NOH-Ap interface. Thus, both the dehydration of the nanolayers and the restructuring of the NOH-Ap interface in the coplanar assemblies are tightly related phenomena.

The experimental OH-Ap-to-mature-bone and OH-Ap-to-young-bone red shifts of the Ca 2p excitations display complex T-dependences. They cannot be evidently described in terms of the phonon broadening as the x-ray diffraction studies [45] display that the related cell volume of OH-Ap and the lattice parameters do not change noticeably up to 900 °C. By analyzing the experimental thermal-induced changes in bone we suggest two main mechanisms of their origin. The first mechanism is related with the dehydration of the inter-crystallite regions and leads to their approaching to empty spaces. Applying the 3DSL model to NOH-Ap separated by empty spaces we expect both the gradual increase of the OH-Ap-tobone red shift and the narrowing of the Ca 2p excitations in bone. The second mechanism is related with the atomic restructuring and changing of chemical bonds in the inter-crystallite space. Both the unfolding of the solvation shells for T > 50-60 °C and the degradation of the collagen matrix for $T > 340 \,^{\circ}C$ initiate new chemical reactions that form non-regular Ca²⁺ bonding sites in mineralized bone. The joint action of the mechanisms causes the irregular thermal behavior of the spectroscopic characteristics presented in tables 1 and 2.

Figure 3 illustrates how the restructuring affects the Ca²⁺ 2p⁶ \rightarrow 2p⁵3d¹(*a*₂) and 2p⁵3d¹(*b*₂) line shapes in bone. Each position (*j*) of Ca²⁺ is characterized by



the surroundings potential $W_{Ca(j)}$. The line shape $F_{A(B)}(\omega)$ of the transitions is obviously a sum over all non-equivalent bonding sites

$$F_{A(B)}(\omega) = \sum_{j \in J(\mathbf{T})} c_j f_j(\omega, \omega_j).$$
(2)

Here c_i and J are the concentration of $Ca^{2+}(j)$ ions and the number of the non-equivalent bonding sites, $h\nu_i$ is the transition energy dependent on the surrounding potential $W_{Ca(j)}$. In bone the maximal number J_{max} of the non-equivalent positions is evidently bigger than in OH-Ap. In addition to the well-known Ca1 and Ca2 positions in OH-Ap (see, e.g. [41, 46]) there are Ca²⁺ ions surrounded by the solvation shells in the hydrated layers. On this background the set J depends on T in (2). The distribution function $f_i(\omega, \omega_i)$ of an individual core-to-valence transition can be described with the Voight profile. The corresponding Lorentzian width specifies the inverse Ca 2p-hole lifetime and Gaussian one is responsible for the bandwidth of the photon source and the phonon broadening in NOH-Ap. For simplicity only two $Ca^{2+}(j)$ and $Ca^{2+}(j')$ positions are included in figure 3. The figure illustrates the nonequivalent contributions of the $Ca^{2+}(j')$ 2p excitations to the absorption lines A and B. Since the energies of the terms a_2 and b_2 vary non-uniformly with changing the surrounding potential W_{Ca} these lines demonstrate different spectral shift and broadening. Thus, new Ca²⁺ bonding sites produce not identical changes in the line shape of the Ca 2p excitations. The spectroscopic data presented in tables 1 and 2 confirm the different behavior of the lines A and B. This mechanism is expected to be dependent on the elemental composition of bone too but this aspect is not discussed here.

The experimental results show that the OH-Apto-bone red shifts and changes in the FWHM of the Ca 2p excitations in the mature bone are more pronounced than in the young bone. To understand this difference, we draw our attention that amorphous calcium phosphate is also present in the mineralized bone [47]. The interrelationship of amorphous to crystalline phases changes rapidly with age. Using x-ray diffraction Termine and Posner [47] have demonstrated that the part of the amorphous material in the mineral composition of rat femur decreases with age. In particular, in a three-day-old bone the percentage of crystalline apatite and amorphous calcium is 32.8% and 67.2%. In a 10- and 80-day-old bone these values are 37.6% and 62.5% as well as 63.1% and 32.9% respectively. So, in less than 80 days, the ratio changes in 4 times. The changing of the amorphous/crystalline mineral composition with increasing age is almost identical to the results obtained by using infrared spectroscopy and electron spin resonance method [48, 49]. The noticeable presence of the amorphous component will weaken the influence of the super periodicity on the Ca 2p excitations of young bone tissue. The experimental data in tables 1 and 2 support this effect. Note the temperature treatment also influences the interrelationship but much weaker. According to the work [47] the air drying of the mature bone samples at different temperatures resulted in changes of the amorphous component. Its contribution is 23.8% (at 25 °C), 29.9% (110 °C) and 28.6% (250 °C).

In the framework of the 'black-nanoboxes-inmuddy-waters' concept [18] the 3DSL model describes the electronic structure of mineralized bone in terms of the electro-optical characteristics of its nanoelements. The thermal-induced dehydration and restructuring can be taken into account by introducing the T-dependent electro-optical thickness \tilde{d} of the inter-crystallite region. Using the theoretical

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results [18] and the experimental data analysis we write the hierarchy induced OH-Ap-to-bone red shift of the $Ca^{2+} 2p \rightarrow 3d$ transitions in mineralized bone as

$$\delta E_{\rm 3d}({\rm T}) \approx 2\gamma({\rm T})E_{\rm 3d}\frac{\tilde{d}({\rm T})}{\langle L\rangle}.$$
 (3)

 $\langle L \rangle$ is the mean size of the NOH-Ap in bone and γ is the fraction of Ca²⁺ ions forming the superlattice to their total number in bone. Equation (3) describes the superlattice effect on the 2p \rightarrow 3d transition energy. It depends linearly on the 3d-band energy in OH-Ap E_{3d} , the coefficient γ and the ratio of the electronoptical thickness \tilde{d} of the spacer layers to the mean linear size of the crystallite. According to [24] the OH-Ap-to-bone spectral shift is not inherent in transitions occurring in Ca²⁺ ions which do not form the superlattice in bone. On this background we assume that the non-regular Ca²⁺ sites in the NOH-Ap interface and the amorphous material do not contribute to $\delta E_{3d}(T)$.

As an example, we use the values $E_{3d} \approx 2 \text{ eV}$ [50], $\gamma \approx 0.8$ [47], $\tilde{d} \leq 2 \text{ nm}$ and $\langle L \rangle = \frac{1}{3}(20 + 7.5 + 3.5) \approx 10 \text{ nm}$ [18], apply (3) and obtain the OH-Ap-to-bone red shift in mature bone $\delta E_{3d} \leq 0.3 \text{ eV}$. Thus, the 3DSL model estimation of the shift is in good agreement with the experimental data in table 1. We also note that the thermalinduced changes of the shift δE_{3d} are determined by the coefficient and the geometric ratio $\tilde{d}/\langle L \rangle$ but its age-related changes are controlled by the γ and the mean size $\langle L \rangle$.

The important role of the dehydration and restructuring mechanisms in the thermal-induced changes is supported by the near O 1s edge NEXAFS [18] and infra-red (IR) spectroscopic studies [51, 50, 52]. The suppression of the lowest O 1s absorption peak located at 532.2 eV and the 3400 cm⁻¹ band in IR spectra of heated bone points unambiguously on the thermal-induced removal of water from the intercrystallite space [18, 50] and the destruction of the OH⁻(H₂O)_m and Mg/Ca²⁺(H₂O)_n clusters [18] that dominate the solvation structure in bone at RT. It is the electronic transitions from O 1s level to the low unoccupied molecular orbitals in the clusters that are regarded as the main origin of the 532.2 eV absorption peak [18]. The broad band in IR spectra is tightly related with O-H vibrations give rise in hydrated layers in bone. The molecular dynamic simulations [53] performed to understand the hydration thermodynamics of Ca²⁺ and other ions indicate that the van-der-Waals interaction forms the solvation shells with the average coordination number 7.3. This number agrees with the experimental values extracted from the x-ray, neutron scattering and EXAFS measurements [53–56]. Thus, the suppression of both the absorption peak at 532.2 eV and the vibration band at 3400 cm^{-1} in heated bone [18, 50, 52] makes evident importance of the dehydration and restructuring mechanisms in the hydrated-layer-NOH-Ap-surface network in bone tissue.

Further experimental and theoretical research of local electronic and atomic structure of bone is required. The probing by using O 1s and P 2p excitations and performing *in situ* x-ray absorption measurement of bone samples and their thermal treatment will permit us to increase the accuracy and to understand in more detail the link of electron and atomic structure with the hierarchical organization of skeleton.

5. Conclusion

The performed high energy resolution Ca 2p NEXAFS measurements of mature and young bones have (i) confirmed the predicted OH-Ap-to-bone red shift of the calcium 3d-states, (ii) revealed the irregular temperature dependence of the Ca 2p excitations of bone tissue and (iii) demonstrated the different thermal dependence of the line shapes and the Ca $2p \rightarrow 3d$ transition energies in mature and young bone. The measured OH-Ap-to-bone red shifts of the transition energy vary irregular in the interval 0.1 eV $\leq \delta E_{3d} \leq 0.3$ eV when T grows from RT up to 450 °C. At RT the shifts are found equal $\delta E_A^m \approx \delta E_B^m \approx 0.2 \text{ eV}$ and $\delta E_A^y \approx \delta E_B^y \approx 0.1 \text{ eV}$ for mature and young bone respectively. The shifts averaged over the heating T interval are 0.18 eV, 0.2 eV and 0.14 eV.

The thermal-induced dehydration of the nanolayers leads to the unfolding of solvation shells of ions in the hydrated layers, which launches new chemical reactions as the dielectric shielding weakens and to the subsequent restructuring of the interface in bone. The 3DSL model gives us a key for understanding the relationship between the hierarchical organization of bone tissue and its electronic and atomic structure, thermodynamics and spectral distribution of oscillator strength for core-to-valence transitions in it.

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