# OBSERVATIONS OF IRISH *PARECHINISCUS* CUÉNOT, 1926 (HETEROTARDIGRADA: ECHINISCIDAE) CONVEY NEW INFORMATION (AND QUESTIONS) ABOUT THE GENUS

# Erica DeMilio, Denis V. Tumanov, Colin Lawton and Jesper Guldberg Hansen

# ABSTRACT

*Parechiniscus* Cuénot, 1926 is a monotypic genus of tardigrades that includes among its characteristic features the uncommon trait of unpaired dorsal cuticular plates at the position of each of the first three leg pairs. Our morphological analyses of Irish *Parechiniscus* specimens, which included scanning electron microscopy, led to a new interpretation of configuration and details of the dorsal cuticular plate morphology in this genus. We also provide comments on *Parechiniscus* morphometry, sensory organs and sexual system. New emendations to the genus diagnosis based on our findings are proposed. Additionally, we record *Echiniscus spiniger* Richters, 1904 and *Testechiniscus spitsbergensis spitsbergensis* (Scourfield, 1897) from Ireland for the first time.

# INTRODUCTION

Tardigrades (phylum Tardigrada Doyère, 1840) are microscopic animals (adults typically 100µm-1,000µm) that periodically moult chitinous cuticles (Greven et al. 2016). The tardigrade cuticle has taxonomically important characteristics (Czerneková and Vinopal 2021) that in many heterotardigrades include the configuration and morphology of dorsal and ventral cuticular plates. Cuénot (1926) described Parechiniscus chitonides based on tardigrade specimens, collected from several French locations, that exhibited unique characteristics including bar-shaped, unpaired dorsal cuticular plates that are weakly developed at the body anterior. Considering the unusual condition of the plates of the specimens as a possible orthogenetic representation between unplated marine species and the plated terrestrial taxa, the new genus Parechiniscus Cuénot, 1926, was erected for the specimens. This preceded the definition and division of the main classes of Tardigrada, Heterotardigrada Marcus, 1927 and Eutardigrada Richters, 1926. Additional morphological details, including observant representations of the dorsal plates of P. chitonides, were given by Thulin (1928), Cuénot (1932) and Marcus (1936), but these were limited by the early stage of tardigrade taxonomy in which they were produced.

A second Parechiniscus species, P. unispinosus da Cuhna, 1947, was described from Portugal. However, this taxon was later regarded as synonymous with Hypechiniscus gladiator (Murray, 1905) (Ramazzotti and Maucci 1983; Fontoura et al. 2009; Gasiorek et al. 2021a). Another taxon, also with bar-shaped unpaired plates, was later collected from Utah, U.S.A. and was originally attributed to Parechiniscus by Schuster (1975). This species was subsequently designated Novechiniscus armadilloides (Schuster, 1975), the type of a new genus by Kristensen (1987) in his important revision of the Echiniscidae. Thus, Kristensen (1987) regarded Parechiniscus as monotypic and revised the genus with emendations based upon specimens identified as 'P. chitonides' from the Thulin collection and fresh specimens collected from Sweden. Kristensen (1987) determined that Parechiniscus and Novechiniscus Kristensen, 1987 were not closely related despite the superficial resemblance in the form of some dorsal plates. This was later supported by the detailed morphological analyses of Novechiniscus involving SEM by Rebecchi et al. (2008) and phylogenetic inferences by Jørgensen (2000) and Gasiorek et al. (2018a). The uniqueness of the *P. chitonides* morphology was emphasised through these works along with its distinct phylogenetic position within Echiniscidae, also inferred via molecular analyses (e.g. Jørgensen et al. 2011; Guil et al. 2013).

Erica DeMilio (corresponding author; email: erica.demilio@ gmail.com. ORCID iD: https://orcid.org/0000-0002-5908-5542), Zoology, School of Natural Sciences, Ryan Institute, National University of Ireland Galway, Galway, Ireland; Denis V. Tumanov, (Independent researcher); Colin Lawton, Zoology, School of Natural Sciences, Ryan Institute, National University of Ireland Galway, Galway, Ireland and Jesper Guldberg Hansen, Natural History Museum of Denmark, University of Copenhagen, Copenhagen, Denmark.

Cite as follows: DeMilio, E., Tumanov, D.V., Lawton, C. and Hansen, J.G. 2022 Observations of Irish Parechiniscus Cuénot, 1926 (Heterotardigrada: Echiniscidae) convey new information (and questions) about the genus. Biology and Environment: Proceedings of the Royal Irish Academy 122B (3).

Received 13 July 2022. Accepted 26 September 2022. Published 3 November 2022.

Parechiniscus was only recently reported from Ireland for the first time (DeMilio et al. 2022). A sample of moss containing specimens identified as Parechiniscus cf. chitonides was collected at Glensleade, County Clare, Republic of Ireland as part of the All-Ireland Tardigrade Survey (AITS). Studies of these specimens conducted in light microscopy (LM) and scanning electron microscopy (SEM) presented the opportunity to contribute new information about Parechiniscus morphology. As part of the present study, additional moss material of Tumanov (2005) was revisited, which was also found to contain P. cf. chitonides as well as two other heterotardigrade taxa, Echiniscus spiniger Richters, 1904 and Testechiniscus spitsbergensis spitsbergensis (Scourfield, 1897). Both of these species represent new records for Ireland (Republic of Ireland and Northern Ireland).

## MATERIALS AND METHODS

Specimens from a population of Parechiniscus cf. chitonides were collected at Glensleade, County Clare, Republic of Ireland in moss on limestone pavement as reported by DeMilio et al. (2022). Mosses were collected from the substrate surface and allowed to dry in paper envelopes for storage. Following a 24-hour rehydration period in distilled water, moss material was agitated vigorously in a glass jar. The contents were passed through a set of sieves of 1.22mm and 36µm diameter. The retained material was inspected under a stereomicroscope at magnifications of at least  $40 \times$ to manually remove tardigrade specimens. Retrieved specimens were either mounted on microscope slides in Heinz's polyvinyl alcohol (PVA) or Hoyer's medium or prepared for scanning electron microscopy (SEM). Specimens intended for SEM analysis were prepared according to the protocol described by Perez-Pech et al. (2020) or Tumanov (2020). Slide mounted specimens were examined with differential interference contrast (DIC) and phase contrast (PhC) up to the highest available magnification (100× oil objective and 2× magnification changer) on an Olympus BX51 or Leica DM2500 microscope. An AMScope T720 with PhC kit was also used to view some specimens. Micrographs were made with a digital camera and AMScope capture software (Version 3.7) or using a Nikon DS-Fi3 digital camera with NIS software. Figures were designed using Corel Draw Graphics Suite X6.

Additional material that had been collected as part of the study by Tumanov (2005) from the vicinity of Bellharbour, Co. Clare, Republic of Ireland was further studied. Taxa that were not previously reported were identified to species level.

Morphometric data were obtained as described by DeMilio *et al.* (2022). Statistical analyses were performed with JASP (Version 0.12.2). The methodology of Bartels *et al.* (2011) was followed to check for evidence of allometric growth in morphological analyses.

# RESULTS

A total of 27 *P.* cf. *chitonides* individuals and a single exuvium containing 2 eggs were extracted from moss collected at Glensleade. Twenty-three individuals were mounted on slides (10 females, 6 males, 5 of undeterminable gender, and 2 four-clawed juveniles) and four individuals were prepared for SEM.

The revisited material of Tumanov (2005) contained previously unreported specimens of *P. cf. chitonides* from the vicinity of Bellharbour from moss on rock (rock type unspecified). A total of 37 *P. cf. chitonides* individuals were extracted. Twenty-seven individuals were mounted on slides and ten individuals were prepared for SEM. Representative specimens of Irish *P. cf. chitonides* are presented in Figures 1–9.

# DESCRIPTION OF IRISH PARECHINISUCUS CF. CHITONIDES

#### Body morphology and morphometry

Cuticle, translucent to light brown before and after mounting in PVA or Hoyer's media, with distinctive sculpture elements (Figs 1, 2) (described below). Body length range 122.9-202.3µm (Glensleade population). Additional measurements of selected structures are given in Table 1 and Table 2. None of the measured characters showed evidence of allometric growth (i.e. when log transformed, the slope (b) of the regression line of the measured trait values relative to body length was not significantly different from a slope of 1 as shown by t-tests (p < 0.05)), and therefore trait measurements were determined to be suitable for use in a morphometric ratio (pcap, see below). The slope (b) of the regression line of each measure trait relative to body length with log-log transformed data is given in Table 1. Although only isometric growth was indicated, the Y-intercept (a\*) of Thorpe's transformed traits are also given in Table 1 for potential comparisons with future studies as recommended by Bartels et al. (2011).

#### Cephalic structures

Black, round eyes, not composed of granules, persist after mounting in PVA. Cephalic structures observed in LM (Fig. 3) and SEM (Fig. 4). Cephalic appendages include small internal and external cirri (Fig. 3B), both without true cirrophores, with slightly swollen bases. External cirri positioned upon an elevated lobe (Fig. 4A). Secondary clava



Fig. 1—Irish *Parechiniscus* cf. *chitonides* habitus (DIC). A female, dorsal; B female, ventral. cxp= coxal plate, fp= femoral plate, p4= leg sense organ IV, filled white arrowhead indicates evident area of large intracuticular pillars.

with narrow base and expanded, laminar paddle shaped distal part (Figs 3; 4A,C). Paired planate, rounded tertiary clavae (Figs 3; 4A,C) present rostral to secondary clavae. Primary clava in the form of an elongated papilla with a blunt apex (Fig. 3C), situated at the cirrophore base of cirrus A (Fig. 3A), a long flexible filament. Other dorsal and lateral sensory appendages absent. Ten peribuccal papulae present, eight fused in pairs and two singles (Fig. 4B). Single papulae positioned along median sagittal plane. Internal buccal-pharyngeal structures not well observed.

#### Dorsal cuticular plates and sculpture elements

Dorsal plates unpaired, distinctly sculptured. Dorsal plates sometimes poorly defined, especially towards the cephalic end (Figs 1, 2). Nine dorsal plates (Figs 5, 6) include: cephalic plate (cep), neck plate (np), three single transverse plates (t), three median plates (m), and caudal plate (cap). Lateral supplementary plates (lsps) also present.

Dorsal sculpture elements (Figs 3C; 4C,D; 7A,B) include: visible intracuticular pillars (some finer and diffuse) that appear as small, raised circular



Fig. 2-Irish Parechiniscus cf. chitonides habitus (SEM). A dorsal; B lateral.



Fig. 3—*Parechiniscus* cf. *chitonides* cephalic structures from various perspectives (DIC). A lateral, B lateral; C dorsal; D ventral. cA= cirrus A, ex= external cirrus, ic= internal cirrus, pc= primary clava, sc= secondary clava, tc= tertiary clava, filled black arrowhead indicates cuticular pore.



Fig. 4—*Parechiniscus* cf. *chitonides* cephalic structures (SEM). A head, lateral; B mouth ring with peribuccal papulae; C head, dorsal; D cephalic plate pores. sc= secondary clava, tc= tertiary clava, filled black arrowhead indicates cuticular pore.

points in DIC or regular, round dark dots in PhC; pores that appear as small pits between pillars in DIC or as minute, light points in PhC; depressions that appear as rounded or irregular, singular or adjoining indentations in DIC or as light areas creating a mottled pattern in PhC; and epicuticular elements (EEs) that appear as evident, rounded bumps in DIC or evident, dark bulky circular or subcircular areas in PhC.

Cephalic plate poorly defined, undivided (Fig. 1A). In LM cep appears sculptured with visible intracuticular pillars on its entirety and diffuse fine pillars surrounding all margins, scattered pores, and large, round, wart-like EEs of varying sizes (Figs 3A,C; 5A). In LM, EEs appear not as well defined on cep as on other dorsal plates. In SEM three sculpture element types well visible on cep including surface indications of pillars, large pores along cep anterior-anterolateral margin (Figs 4C,D) with additional pores at the plate interior, and wart-like EEs equally as well developed as on other dorsal plates (Fig. 2B). Depressions absent from cep. Cep terminates caudally at a shallow crevice delimiting the cephalic region (Figs 2, 4C). Lateral cephalic plate absent but corresponding area clearly sculptured with visible intracuticular pillars and surrounded by surface devoid of visible pillars (Figs 3A,B).

Neck plate broad, without divisions, sometimes with poorly defined margins (Figs 1A; 3A,C; 5A). Cirrus A and primary clavae insertion positions encompassed by np (Figs 2A; 5; 6A,B). Np sculptured with visible intracuticular pillars, scarce pores, and wart-like EEs on its entirety (Figs 2; 3A,C; 5; 6A,B). Depressions absent. In LM, border between cep and np may be difficult to discern more evident in SEM. Np terminates caudally at a crevice in which EEs are absent or very sparse (Figs 2; 5B; 6A,B).

Transverse plate 1 (t1), unpaired, bar-shaped with lateral broadenings, positioned at level of legs I, evidently narrower than np. T1 sculptured with visible intracuticular pillars, scarce pores, and large,

# BIOLOGY AND ENVIRONMENT



Fig. 5—Colourisation demonstrating *Parechiniscus* dorsal cuticular plate configuration. A lateral (DIC); B lateral (SEM). cep= cephalic plate, np= neck plate, t= transverse plate, m= median plate (a= anterior part, p= posterior part), cap= caudal plate.

densely distributed wart-like EEs on entire plate (Figs 1A; 2; 5; 6A,B). Depressions absent and median transverse groove absent from t1. Transverse plate 2 (t2) positioned slightly anteriorly to level of legs II, formed as an anterior and posterior latitudinal ridge separated by a broad median transverse groove that is interrupted at its median by a longitudinal line of EEs upon a ridge (sometimes incomplete) (Figs 2; 5; 6A,C). Epicuticular elements may be present within median transverse groove. Larger, more densely distributed EEs occur on both anterior and posterior t2 ridges than within median groove. Visible



Fig. 6—Colourisation demonstrating *Parechiniscus* dorsal cuticular plate configuration (SEM). A uncoloured overview, whole animal; B anterior body region; C middle body region; D posterior body region. cep= cephalic plate, np= neck plate, t= transverse plate, m= median plate (a= anterior part, p= posterior part), cap= caudal plate, long-dashed line indicates median transverse furrow, short-dashed line indicates fold, asterisk indicate lateral supplementary plate.

intracuticular pillars present, more evident on ridges than within median groove. Pores scattered on t2 anterior and posterior ridges. Depressions not observed on t2. T3 positioned slightly anterior to level of legs III, similar in morphology to t2 (Figs 5; 6A,D) but with conspicuous depressions present at the base of its anterior ridge (Figs 2B; 6D). Posterior ridge of t3 with EEs more poorly developed than EEs on posterior ridge of t2 (Figs 2B; 6C,D).

Median plate 1 positioned between t1 and t2. M2 positioned between t2 and t3. M3 positioned between t3 and caudal plate. M1 and m2 share a similar morphology. M1 and m2 each divided into a larger anterior (a) part and smaller inconspicuous posterior (p) part (Figs 5; 6B,C). M1a and m2a lenticular in form and each composed of heavily sculptured anterior and posterior ridges separated by a deep median transverse furrow (Figs 2; 5; 6B,C). Dense visible intracuticular pillars, cuticular pores and largest wart-like EEs occur on m1a and m2a ridges, especially developed on anterior ridges (Figs 1; 2; 5B; 6A–C). Small depressions present at m1a anterior ridge and m2a bases, anterior to the median transverse furrows (Fig. 6C). Depressions not observed on m1a or m2a posterior ridges. M1a and m2a both divided from smaller, narrower, m1p and m2p at a crest (Figs 2; 5). M1p and m2p with intracuticular pillars, mostly smooth but sparse pores



Fig. 7—*Parechiniscus* cuticle. A cuticular sculpture elements (DIC); B cuticular sculpture elements (PhC); C exuvium containing two eggs (DIC). white filled arrowhead indicates intracuticular pillar, white hollow arrowhead indicates depression, black filled arrowhead indicates cuticular pore, black hollow arrowhead indicates epicuticular element (EE).

and small EEs may be present (Figs 6B,C). Small, difficult to observe depressions present on m1p and m2p anterior regions. M1p and m2p each with a median fold (Figs 6B,C). M1p and m2p posterior margins occur at a shared crest upon t2 and t3 anterior ridges respectively (Figs 2; 5). M3 undivided (i.e. without a posterior portion), with a similar morphology to m1a and m2a (Figs 1A; 2B; 5; 6A,D), but with conspicuous and densely distributed depressions present along m3a anterior margin (Figs 1A; 2B; 5A; 6A,D; 7A,B) and sparsely within its transverse median furrow (Fig. 6D).

Caudal plate ovoid. Cap anterior and posterior margins formed as ridges heavily sculptured with large wart-like EEs, pores and visible intracuticular pillars. All three types of sculpture elements present in cap interior along with numerous large, evident, depressions producing an unlevel surface (Figs 1A; 2B; 5; 6A,D; 7A,B). At dorsal plate lateral margins fine cuticular pillars are visible in diffuse areas extending along the lateral body sides (Fig. 7A).

Wart-like EEs, similar to those of dorsal plates present in a network of lines along lateral body sides. Lateral EE lines particularly developed and dense in arcs around areas of insertion of legs I–III to body (Figs 1A; 2B; 5; 6; 7B). Lateral supplementary plates (lsp) present, weakly demarcated and difficult to observe (LM (Figs 1A; 5A) and SEM (Figs 2; 5B), but with clusters of large wart-like EEs (Figs 6A–C). Lsps positioned at either lateral side of m1a and m1p.

#### Leg morphology and leg sense organs

Legs with weakly demarcated coxal and femoral plates (Fig. 1A). Coxal and femoral plates sculptured with wart-like EEs (Figs 2B, 6B,D, 8C). Pores observed on femoral plates (Figs 8A, B). Tarsus well differentiated and sculptured on ventral side with



Fig. 8—Parechiniscus cf. chitonides leg structures. A leg II and claws (DIC); B leg IV (DIC); C leg I and claws (SEM). p4= leg sense organ IV, white filled arrowhead indicates intracuticular pillar, solid line circle indicates area of visible fine pillars, black filled arrowhead indicates cuticular pore, black hollow arrowhead indicates epicuticular element (EE).

intracuticular pillars (Fig. 9B). Dense intracuticular pillars visible elsewhere on leg surfaces (Fig. 8). Leg sense organs I–III absent (Fig. 8C). Large, rounded cone-shaped, sense organ evident on leg IV (p4) positioned between coxal and femoral plates on external dorsal leg surface (Figs 1A; 8B). Claws well developed (Figs 8A,C). External claws smooth. Internal claws with robust downward curving spur positioned distantly from claw base, near to but more proximally than the midpoint of the main branch.

#### Ventral structures

Ventral body surface almost entirely covered with dense and clearly visible intracuticular pillars in LM (Figs 3D; 8A; 9A,B).Visible pillars appear larger than surrounding fine pillars at several positions on ventral body surface (subcephalic area, between each leg pair, area surrounding gonopore and anus, and particularly evident in line with each leg pair I–III (Fig. 1B)). Large, wart-like EEs present in a band composed of several rows (hardly visible in LM, well-visible in SEM (Figs 9C,D) located between the female and male gonopores and anus.

Female gonopore (Figs 9A,C) of six-petaled rosette shape, large, unsculptured, but surrounded on all sides by visible dense intracuticular pillars. Male gonopore visible as a pore within a round shallow, unsculptured depression in LM surrounded by dense intracuticular pillars (Fig. 9B), smaller in diameter than female gonopore ( $\mu$ m and *pcap*, Table 1). In SEM the central thickened structure upon the male gonopore aperture is visible (Fig. 9D). Anal system (Fig. 9 and presented in detail in DeMilio *et al.* 2022) with two rectangular lateral lobes and a fusiform terminal lobe, all lobes well differentiated and clearly observable in LM.

#### Reproduction and developmental stages

Sexual dimorphism: female longer body length than male (Table 1), mean female gonopore to anus distance longer than male (DeMilio *et al.* 2022).



Fig. 9—*Parechiniscus* cf. *chitonides* ventral structures. A female gonopore and anus (DIC); B male gonopore and anus, insert with focus on male gonopore (DIC); C female gonopore and anus (SEM); D male gonopore and anus (SEM). filled arrow indicates gonopore, hollow arrow indicates sculptured tarsus.

Ontogeny: two smooth eggs ( $48.5\mu$ m and  $51.2\mu$ m) observed within an exuvium (Fig. 7C). No two-clawed *Parechiniscus* larvae collected. Two four-clawed juveniles collected (measurements in Table 2) with anal system developed as in adults.

#### NEW IRISH SPECIES RECORDS

The moss material of Tumanov (2005), also collected from County Clare, contained previously unreported tardigrade taxa. In addition to a second population of *P. cf. chitonides* from the vicinity of Bellharbour, specimens of *Echiniscus spiniger* Richters, 1904 and *Testechiniscus spitsbergensis spitsbergensis* (Scourfield, 1897) were obtained (Fig. 10). These taxa represent new species records and the first report of the genus *Testechiniscus* Kristensen, 1987 from Ireland (Republic of Ireland and Northern Ireland).

#### Echiniscus spiniger

Six specimens (adult females) were extracted from a moss sample collected at the same location as the Bellharbour sample containing P. cf. chitonides. E. spiniger belongs to the 'spinulosus group' and undoubtedly requires redescription based on material from the type location (Miller et al. 2010; Gasiorek and Degma 2018; Gasiorek and Michalczyk 2020; see also Pilato et al. 2005). The Irish specimens conform to previous descriptions from other locations in several distinguishing characters. The Irish E. spiniger (Fig. 10A) is similar to that of Maucci (1986) in the dorsal cuticular plate sculpture element morphology (Fig. 10C) and distribution (e.g. paired plates with a band of poorly developed sculpture elements at the anterior margin) and dorsal and lateral appendage morphology (e.g. laterally and dorsally appendages in positions B-D are spinous with the dorsal spines at C<sup>d</sup> longer than those



Fig. 10—Species representing new records for Ireland. A Echiniscus spiniger Richters, 1904 habitus (PhC); B Testechiniscus spitsbergensis spitsbergensis (Scourfield, 1897) habitus (PhC); C E. spiniger scapular plate cuticular sculpture elements (DIC); D T. s. spitsbergensis scapular plate cuticular sculpture elements (DIC).

at D<sup>d</sup> (C =  $33.0\mu$ m, C<sup>d</sup> =  $58.3\mu$ m, D =  $25.9\mu$ m, D<sup>d</sup> =  $43.4\mu$ m, as measured on the best-preserved specimen). See also Pilato *et al.* (2008), Gąsiorek and Degma (2018) and Gąsiorek and Michalczyk (2020) for further details on the comparative morphology of the species.

#### Testechiniscus spitsbergensis spitsbergensis

A single specimen (adult female) was extracted from the same sample with P cf. *chitonides*. The morphology of this specimen (Fig. 10B) conforms to the modern redescription of the nominative subspecies by Gąsiorek *et al.* (2018b), representing a morph with the genus typical cuticular sculpture (Fig. 10D) and the D<sup>d</sup> appendages in the form of a short spine and at least two small spines at the caudal plate posterior margin. Currently, reliable records of this subspecies indicate a distribution confined to the Western Palearctic only (but see discussions in Dastych 1973; Gąsiorek *et al.* 2018b).

# DISCUSSION

#### IRISH PARECHINISCUS CF. CHITONIDES

Observations in LM and SEM of Irish *Pare-chiniscus* cf. *chitonides* specimens yielded new information about the dorsal plate configuration, cuticular sculpture elements, leg plates, sensory organs (cephalic and leg sense organs) and reproductive structures.

lensleade population). cap= caudal plate,	<sup>+</sup> see DeMilio et al. (2022) for descriptive	
alues for selected characters of adult Irish Parechiniscus cf. chitonides specimens (Gle	eight, $\emptyset$ = diameter; $\overline{x}$ = mean, SD= standard deviation, b= slope, a* =Y-intercept. <sup>+</sup>	
able 1—Morphometric v	= length, w= width, h= h	tatistics by sex.

		Femal	es		Male	ş			X	S	D		
							- 4 -					Ļ	ۍ * د
Character	n	шц	pcap	u	m	pcap	1	шц	pcap	шц	pcap	)	3
body length	6	139.3–202.3	479.2-650.5	9	126.1–171.5	475.3-684.7	15	160.8	561.1	18.7	59.0	ı	I
cap	6	28.4-32.7	ı	9	24.8-28.3	ı	15	28.7	I	2.3	ı	0.35	28.56
internal cirrus	$\infty$	4.2 - 6.0	14.5 - 19.6	S	4.5 - 5.6	17.0 - 21.6	13	5.1	18.0	0.5	1.9	0.62	5.34
external cirrus	6	6.9-8.6	23.6-28.5	Ś	6.1 - 9.2	23.3–37.2	14	7.6	26.6	0.8	3.4	0.63	7.71
cirrus A	6	27.1 - 32.6	87.2-105.5	9	25.8 - 31.0	97.9–124.8	15	29.0	101.4	2.0	8.2	0.24	29.51
cirrus A $pbl$	6	13.4-	20.6%	9	17.2%	-20.6%		18.	.2%	1	6.	ı	ı
primary clava	2	3.2 - 5.0	11.0 - 15.4	9	3.5-4.7	12.4–18.8	13	4.1	14.3	0.6	1.9	0.46	4.13
secondary clava l	9	5.6-8.5	19.3 - 28.0	ς	6.3-7.4	22.5-26.2	6	7.1	24.3	1.0	27	0.73	7.16
secondary clava w	9	4.1 - 5.0	13.7 - 16.3	m	4.5-5.2	16.0 - 18.9	6	4.6	15.9	4.1	13.7	0.13	4.81
external claw I h	6	6.6-7.7	21.7 - 24.9	9	6.3-7.2	23.5 - 29.1	15	6.9	24.2	0.3	1.7	0.18	6.90
internal claw I h	6	7.3-8.2	23.2–27.1	9	6.9–7.3	25.0–29.6	15	7.5	26.1	0.4	1.5	0.28	7.44
spur I l	6	1.5 - 2.1	5.0-7.2	9	1.4 - 1.8	5.5-6.5	15	1.7	5.9	0.2	0.5	0.34	1.70
spur I insertion point	6	1.8 - 2.4	5.9 - 8.3	9	1.7 - 2.3	7.0-8.1	15	2.1	7.4	0.2	0.7	0.19	2.09
external claw IV h	2	7.7-8.9	25.6–29.4	c	8.0-8.5	28.2–34.3	10	8.3	28.4	0.3	2.5	0.10	8.28
internal claw IV h	9	8.5–9.9	27.3–31.4	m	8.9–9.1	31.5–36.8	6	9.1	31.2	0.4	2.6	0.08	9.06
spur IV 1	9	1.8 - 2.6	6.0-8.3	m	1.9–2.4	6.6–9.9	6	2.2	7.5	0.3	1.2	0.74	2.16
spur IV insertion point	9	2.9 - 3.6	9.4–11.5	c	3.1 - 3.4	10.9 - 12.4	6	3.1	10.7	0.2	1.1	0.04	3.19
sense organ leg IV	2	2.6 - 3.4	9.2 - 10.9	9	2.3 - 3.3	8.7-13.0	13	2.9	10.4	0.3	1.2	-0.12	2.88
gonopore $\omega^+$	2	8.8 - 10.6	26.9–35.5	9	2.2-3.5	7.7–12.6	ı	ı	ı	ı	·	ı	ı
gonopore to anus <sup>+</sup>	$\infty$	15.3-22.6	51.3-79.1	9	13.7–17.1	51.9-67.7	ı	ı	I	ı	,	ı	ı
gonopore to anus $pbl^+$	8	10.1%	-13.4%		9.3-	11.6%	ı		I			ı	ı

	Juveniles					
	n -	x		S	D	
Character		μm	рсар	μm	pcap	
body length	2	128.9	608.2.1	8.4	116.6	
cap	2	21.5	-	2.8	-	
internal cirrus	2	3.6	16.8	0.3	0.8	
external cirrus	2	7.6	26.6	0.8	3.4	
cirrus A	2	21.7	102.5	2.3	23.6	
cirrus A <i>pbl</i>	2	16.8% 0.7		.7		
primary clava	2	2.6	12.4	0.1	2.0	
secondary clava l	0	-	-	-	-	
secondary clava w	0	-	-	-	-	
external claw I h	2	5.2	24.5	0.3	4.3	
internal claw I h	2	6.2	29.1	0.6	6.9	
spur I l	2	1.1	5.3	0.1	1.2	
spur I insertion point	2	1.6	7.5	0.1	1.6	
external claw IV h	2	6.4	29.9	0.1	3.3	
internal claw IV h	2	7.0	32.6	0.1	4.0	
spur IV l	2	1.3	6.2	0.1	1.7	
spur IV insertion point	2	1.9	8.8	0.3	0.3	
sense organ leg IV	2	1.9	8.5	0.2	0.3	

Table 2—Mean morphometric values for selected characters of four-clawed juvenile Irish *Parechiniscus* cf. *chitonides* specimens (Glensleade population). cap=caudal plate, l=length, w=width, h=height,  $\bar{x}$ =mean, SD=standard deviation.

#### Characterisation of the dorsal cuticular plates

The conformation and the morphology of the dorsal cuticular plates of *Parechiniscus* is unique among Echiniscidae. We therefore proposed a dorsal plate terminology slightly modified from that in general use for other echiniscids. Our LM and SEM observations found that nine dorsal cuticular plates are present: cephalic plate (cep), neck plate (np), three single transverse plates (t), three median plates (m) and caudal plate (cap). Weakly developed lateral supplementary plates (lsp) are also present.

Cuénot (1926) did not specify the number of plates present in the original description of P. chitonides. Thulin (1928) gave a detailed account of the dorsal morphology, which he regarded as a system of plates, crests and furrows with associated musculature. He was however, limited to what could be seen in LM, and so could not discern the anterior most weakly developed plates, or the precise divisions between more posterior plates, as were observed in our analyses. In his revision, Kristensen (1987) determined the presence of 8-9 dorsal plates and illustrated their general form (Fig. 36 in Kristensen 1987) differently to Cuénot (1926) and Thulin (1928). Although the number of dorsal plates we identified on the Irish specimens overlaps with Kristensen's (1987) count, we concluded a different plate configuration to each of these earlier authors. The flexibility of the ridge-shaped dorsal plates when

subjected to coverslip pressure for slide mounting in LM and the use of SEM in the present study likely account for these differences.

Cuénot (1926), Thulin (1928) and Kristensen (1987) all commented on the poor definition of the anteriormost plates. The cep and np were not recognised by Thulin (1928) nor Kristensen (1987) due to the weak sclerotisation of these plates, although their borders are recognisable with modern LM and in SEM. The *Parechiniscus* np is much broader than in other echiniscid genera in which it typically occurs in the form of a narrow bar, sometimes indiscernible from the adjacent plates in LM (e.g. compare to np of *Hypechiniscus* Thulin, 1928 (Kristensen 1987; Gąsiorek *et al.* 2021a) *Pseudechiniscus* Thulin, 1911 (Kristensen 1987; Tumanov 2020) and *Testechiniscus* (Gąsiorek *et al.* 2018b)).

Another difference in dorsal plate morphology is that *Parechiniscus* deviates from the condition of most echiniscid genera, which typically have a scapular plate (scp) and paired plates (sometimes considered as 'segmental plates' (Kristensen 1987)) located at the level of each of the first three leg pairs. In most other echiniscid genera the scp is typically positioned dorsal to legs I and extends at its anterior margin to the posterior of the primary clava and cirrus A. The scp can often be well defined and strongly sclerotised (e.g. in the genera *Bryodelphax* Thulin, 1911(DeMilio *et al.* 2022), *Echiniscus* Schultze, 1840 (Kristensen 1987; Gąsiorek *et al.* 2017) and *Mopsechiniscus* (Guidetti *et al.* 2014)). In other genera it is common for the scp to be poorly sclerotised and demarcated (e.g. *Hypechiniscus* (Kristensen 1987; Gąsiorek *et al.* 2021a) and *Pseudechiniscus* (Kristensen 1987; Tumanov 2020)). The scp has had importance in the use of its measurement as the basis for the *psc* value, the standard per cent ratio between the length of a structure and the scapular plate length (Dastych 1999; Fontoura and Morais 2011) widely used in echiniscid comparative morphometrics.

In Parechiniscus, our observations led to a new interpretation of the dorsal plates that correspond to the scp and paired plates in most other echiniscid genera. These plates are positioned slightly anterior to each the first three leg pairs, are all singular and have a general shape of a transversally oriented bar, therefore we refer to them as transverse plates (t1t3). The interplate borders between the transverse plates and the adjacent plates are sometimes poorly discernible in LM. We first attempted to utilise t1 in a morphometric ratio in a similar way to the scp of other genera but found that in LM the margins of t1 were often not sufficiently defined as to be able to obtain a reliable measurement. Therefore, we used the strongly sclerotised caudal plate (cap) measurement (µm) to calculate a morphometric index, *pcap*, which we define as the per cent ratio between the length of a structure and the caudal plate length. Although only a small sample size was available, we did not find evidence of allometric growth in Parechiniscus caudal plate compared to body length calculations. We suggest the use of the pcap ratio in place of the psc ratio for Parechiniscus to reduce body size effect in morphometric comparisons of structures showing isometric growth (Bartels et al. 2011). If future morphometric analyses involving larger sample sizes find evidence of allometry in Parechiniscus structures, Thorpe's normalisation techniques may be used as discussed in the context of tardigrades by Bartels et al. (2011) to eliminate body size effects in comparisons between populations.

Thulin (1928) distinguished, what he termed annular plates ('annularplatten') at the approximate positions of t2 and t3. Kristensen (1987) figured segmental plates II and III at similar locations. The precise interplate borders and individual plate morphologies as interpreted by these earlier authors are not entirely clear from their illustrations. In our attempt to characterise the Parechiniscus dorsal plates, we found that SEM was critical to understanding the transverse plate morphology, as the actual forms of these plates may be considered counterintuitive to how they can appear in LM. Only in SEM can t2 and t3 be clearly seen to have structure composed of a double ridge separated by a median transverse groove, homologous in appearance to the paired plates of other echiniscid genera.

Similarly, SEM enabled us to observe the form of all three median plates in greater detail than was possible for previous authors. Only Kristensen (1987) recognised the general divided condition of median plates 1 and 2 in LM, but SEM enabled us to more precisely distinguish and characterise the crests and furrows associated with the median plates in Parechiniscus. We only found a single previously published SEM image of an Italian Parechiniscus (Fig. 111 in Maucci 1986), but we were not able to make detailed comparisons with this image due to its low resolution. Similarly, we lacked material from the type population for comparative studies to determine the range of diversity of dorsal cuticular sculpture elements in Parechiniscus. However, based upon our observations and our inferences from previous reports (e.g. Cuénot 1926; Marcus 1936; Arcidiacono 1964; Kristensen 1987) large, wart-like EEs may be a characteristic component of the Parechiniscus dorsal cuticular sculpture.

#### Sense organs

The presence of tertiary clavae has not been previously reported for Parechiniscus. The occurrence of tertiary clavae within each of the two heterotardigrade orders, Echiniscoidea Richters, 1926 and Arthrotardigrada, Marcus 1927, is uncommon. In Echiniscoidea tertiary clavae have been previously reported only in the echiniscid genus Cornechiniscus Maucci and Ramazzotti, 1981 (Kristensen 1987), and according to Gasiorek et al. (2019b) to occur 'occasionally' in Nebularmis Gasiorek and Michalczyk, 2019 in Gasiorek et al. 2019a. The tertiary clavae reported in these other genera differ in form and/orposition to those that we observed on Parechiniscus. Kristensen (1987) showed small dome-shaped tertiary clavae located near the base of the internal cirri in specimens identified as Cornechiniscus cf. holmeni (Petersen, 1951). The tertiary clavae shown on N. reticulatus (Murray, 1905) by Gasiorek et al. (2019b) are papillate and positioned at the level of, but caudal to, the internal cirri. In comparison, on our Parechiniscus specimens we observed considerably large planate, rounded tertiary clavae that were positioned more rostrally in relation the internal cirri and other cephalic sense organs than in the other genera.

While it does not meet modern taxonomic standards, in our opinion, the original description of *Pseudechiniscus megacephalus* Mihelčič, 1951 (*nomen dubium* according to Dastych 2015 and Gąsiorek *et al.* 2021b), might suggest the presence of tertiary clavae (see Dastych 1993 on the status of the Mihelčič collection). Although the identification of the described cephalic structures as potential tertiary clavae was not stated by Grobys *et al.* (2019), we agree with those authors that the presence of such sensory structures should place the taxon in another genus. The thorough re-examination of

Novechiniscus by Rebecchi et al. (2008) involving SEM imaging provided new information on the cephalic sense organs of that taxon. In addition to the dome-like structure upon which Kristensen (1987) reported the internal cirrus to sit, Rebecchi et al. (2008) suggested the presence of an additional dome at the base of the external cirrus. We interpret the published *Novechiniscus* micrographs (Fig. 3A (LM) and Fig. 4A (SEM) in Rebecchi et al. 2008) to suggest that the dome structure reported as constituting the base of the internal cirrus may actually be a potential tertiary clava positioned adjacent to and arching around the internal cirrus.

In Arthrotardigrada, tertiary clavae were reported in the original descriptions of Renaudarctus Kristensen and Higgins, 1984, Paradoxipus Kristensen and Higgins, 1989, Neoarctus de Zio Grimaldi, D'Addabbo Gallo and Morone De Lucia, 1992, and in Angursa Pollock, 1979 by later authors (Noda 1985; Bussau 1992; Fujimoto and Hansen 2019). Tertiary clavae fused to secondary clavae have also been suggested to occur in Coronarctus Renaud-Mornant, 1974 (Hansen et al. 2012) and in Nodarctus Fujimoto and Yamasaki, 2017. The tertiary clavae are lamellar in form in all arthrotardigrade genera in which they are known to occur, except for *Neoarctus* in which they are papillate. The extent of variation in the morphology of tertiary clavae and their position in relation to other cephalic structures in both Arthrotardigrada and Echiniscoidea may not be fully known, as suggested by the new recognition of these structures in Parechiniscus.

The original description of Parechiniscus chitonides stated the presence of a sense organ at the base of leg I in the form of a very small, difficult to observe spine (Cuénot 1926). In the original drawing it appeared coxally positioned but in a later illustration, the leg I sense organ is shown in the femoral region (Cuénot 1932). Kristensen (1987) also reported a small, pointed sense organ on leg I of his Swedish specimens but in his illustration (Fig 36. in Kristensen 1987) suggests a femoral position different to that indicated by Cuénot (1926). We observed some Irish Parechiniscus specimens with a well-developed EE in the femoral region of leg I that could potentially be mistaken for a sense organ in LM. However, our SEM analyses confirmed that the Irish Parechiniscus lack a leg I sense organ. The absence of this structure, specifically stated to be present in the original description of the taxon, required us to use open nomenclature in the identification of the Irish specimens. P. chitonides apparently lacking a leg I sense organ were also reported from Sicily (Arcidiacono 1964). In other echiniscid genera, the presence/absence and morphology of leg sense organs is commonly utilised as a species level character (e.g. Barbaria Michalczyk, Gąsiorek, Morek and Stec, 2019 in Gasiorek et al. 2019a

(Gąsiorek et al. 2022); Bryodelphax (DeMilio et al. 2022); Claxtonia Gąsiorek and Michalczyk, 2019 in Gąsiorek et al. 2019a (Degma et al. 2021); Pseudechiniscus (Tumanov 2020) and Testechiniscus (Gąsiorek et al. 2018b)). The reported variability of the leg I sense organ and any value of this character in discriminating potential species of Parechiniscus warrants further investigation.

#### Sexual system

Both females and males were extracted from the Irish P. chitonides population. Our sample had a sex ratio approximately 2:1, suggesting that males were not uncommon among the Irish Parechiniscus. This is in contrast to the samples of Kristensen (1987) who reported that he collected several hundred P. chitonides specimens from moss on rocks in Sweden but stated that males were not found. This stark difference in the sex composition of these populations could be the result of many factors including the unknown influence of environmental variables, chance of sampling, or a representation of an actual difference in sexual system of two distinct taxa yet to be delineated. The uncertainty of the cause of discrepancy in the occurrence of sexes between two minimally sampled populations should prohibit premature conclusions about the sexual systems or reproductive mode(s) of tardigrade species.

# NEW RECORDS FOR IRELAND

The two echiniscid species found during the present reinvestigation of Tumanov's (2005) material, *E. spiniger* and *T. spitsbergensis spitsbergensis* increase the number of records of Irish heterotardigrades to 22 species in 9 genera (DeMilio *et al.* 2016; DeMilio and Lawton 2020; DeMilio *et al.* 2022). However, the current numbers of recorded tardigrade taxa should not be considered as the final indicators of the extent of the phylum's biodiversity in Ireland. Historically, faunistic surveys involving tardigrades have been rare and highly localised (DeMilio *et al.* 2016), but the number of Irish taxa will certainly increase as further results from other AITS samples are published.

# CONCLUSION

Parechiniscus chitonides was described at a relatively early stage of tardigrade taxonomy at a time when modern imaging techniques and molecular analyses did not exist. Cuénot (1926) did include in his original description a few uncommonly specific details on the morphology and distribution of some sculpture elements on the dorsal and lateral body sides and a clearly stated presence and position of a sense organ on leg I. Despite this and the later attempts to better characterise the species, many traits that are now recognised as important for separating heterotardigrade species and genera have remained unreported for Parechiniscus (e.g. morphology of the leg plates, ventral cuticle sculpture pattern, and anal system). The missing information about these and other taxonomically important characters from other populations and a lack of modern re-description of type specimens make it difficult to determine if records of 'Parechiniscus chitonides' from a broad swath of Europe and North Africa (McInnes 1994; McInnes et al. 2017 and references within) all truly represent a single taxon, or if unrecognised hidden species diversity is present.

A new redescription involving the type material (if it still exists) from one of the original French locations or a designated neotype will provide a baseline for the comparison of specimens from other populations including the Irish population described herein. The integrative re-description of Pseudechiniscus suillus (Ehrenberg, 1853) by Grobys et al. (2020) together with integrative analyses by Cesari et al. (2020) and the comprehensive review of morphological characters by Tumanov (2020) catalysed the delimitation of several new species morphologically similar to Pseudechiniscus suillus that were previously impossible to separate (e.g. Roszkowska et al. 2020; Kayastha et al. 2020). Such research focus on Parechiniscus chitonides might produce a similar outcome. For now, our observations on the Irish Parechiniscus specimens provide new data on several characters. We therefore suggest that this new information justifies that the genus diagnosis is further emended from Cuénot (1926) and Kristensen (1987) as follows:

## PARECHINISCUS CUÉNOT, 1926 EMENDED DIAGNOSIS

Small, black eyes. Cephalic structures include three pairs of clavae. Tertiary clavae planar in form. Rigid buccal tube. Nine unpaired dorsal plates present (cephalic plate, neck plate, three single transverse plates, three median plates and caudal plate).

Composition: Parechiniscus chitonides Cuénot, 1926

Furthermore, the heterotardigrades *Echiniscus spiniger* and *Testechiniscus spitsbergensis spitsbergensis* are added to the checklist of species known from Ireland.

#### FUNDING

#### ACKNOWLEDGEMENTS

The authors thank Vadim Panov for collecting a part of the material studied herein and Dr Dan Minchin for assistance in obtaining additional sampling station data. Drs Thomas Pape and Martin Vinther Sørensen and Charlotte Hansen kindly helped to facilitate our work at the Zoological Museum, Natural History Museum of Denmark. We are grateful for the anonymous constructive comments received during the review process that helped to enhance our manuscript. We thank the Royal Irish Academy editorial staff that handled and improved our submission.

#### REFERENCES

- Arcidiacono, R. 1964 Secondo contributo alla conoscenza dei Tardigradi dei Monti Nebrodi. Bolletino delle Sedute dell'Accademia Gioenia di Scienze Naturali, Catania, S. IV 8, 87–203.
- Bartels, PJ., Nelson, D.R. and Exline, R.P. 2011 Allometry and the removal of body size effects in the morphometric analysis of tardigrades. *Journal of Zoological Systematics and Evolutionary Research* 49, 17–25. https:// doi.org/10.1111/j.1439-0469.2010.00593.x
- Bussau C. 1992 New deep-sea Tardigrada (Arthrotardigrada, Halechiniscidae) from manganese nodule area of the eastern South Pacific. *Zoologica Scripta* **21**, 79–91. https://doi.org/10.1111/j.1463-6409.1992. tb00311.x
- Cesari, M., Montanari, M., Kristensen, R.M., Bertolani, R., Guidetti, R. and Rebecchi, L. 2020 An integrated study of the biodiversity within the *Pseudechiniscus* suillus-facettalis group (Heterotardigrada: Echiniscidae). Zoological Journal of the Linnean Society 188, 717–32. https://doi.org/10.1093/zoolinnean/zlz045
- Cuénot, L. 1926 Description d'un tardigrade nouveau de la faune francaise. *Comptes Rendus de l'Académie des Sciences, Paris* **182**, 744–45.
- Cuénot, L. 1932 Tardigrades. In P. Lechevalier (ed.), Faune de France 24, 1–96. Paris.
- Czerneková, M. and Vinopal, S. 2021 The tardigrade cuticle. *Limnological Review* **21**, 127–46. https://doi. org/10.2478/limre-2021-0012
- da Cunha, A.X. 1947 Description d'un Tardigrade nouveaux de la faune portuguaise. Parechiniscus unispinosus sp. n. Memórias e Estudos do Museu Zoológico da Universidade de Coimbra 180, 1–6.
- Dastych, H. 1973 Systematic studies on Tardigrada, II. Echiniscus clavisetosus Mihelcic 1958, a synonym of Echiniscus spitzbergensis Scourfield 1897. Bulletin de la Société des Amis des Sciences et des Letteres de Poznan, Serie D 14, 89–99.
- Dastych, H. 1993 Redescription of the cryoconital tardigrade *Hypsibius klebelsbergi* Mihelčič, 1959, with notes on the microslide collection of the late Dr. F. Mihelčič (Tardigrada). Veröffentlichungen des Tiroler Landesmuseums Ferdinandeum **73**, 5–12.
- Dastych, H. 1999 A new species of the genus *Mopsechiniscus* Du Bois-Reymond Marcus, 1944 (Tardigrada)

E.D. is the recipient of a postgraduate research scholarship from the Irish Research Council.

from the Venezuelan Andes. *Acta Biologica Benrodis* **10**, 91–101.

- Dastych, H. 2015 Tardigrada. In R. Schuster (ed.), *Checklisten der Fauna Österreichs*, 8, 1–25. Austrian Academy of Sciences Press.
- Degma, P., Meyer, H.A. and Hinton, J.G. 2021 Claxtonia goni, a new species of Tardigrada (Heterotardigrada, Echiniscidae) from the island of Maui (Hawaiian Islands, U.S.A., North Pacific Ocean), with notes to the genus Claxtonia Gąsiorek and Michalczyk, 2019. Zootaxa 4933, 527–42. https://doi.org/10.11646/ zootaxa.4933.4.4
- DeMilio, E. and Lawton, C. Updates to the Checklist of Irish Tardigrada Species (2016–2020, second Edition). https://www.irishtardigrades.com/species-checklists.html. Accessed 26–06–2022.
- DeMilio E, Lawton, C. and Marley N.J. 2016 Tardigrada of Ireland: a review of records and an updated checklist of species including a new addition to the Irish fauna. *ZooKeys* 616, 77–101. https://doi.org/10.3897/ zookeys.616.8222
- DeMilio, E., Tumanov, D.V., Lawton, C., Kristensen, R.M. and Hansen, J.G. 2022 Two new Bryodelphax Thulin, 1928 species (Heterotardigrada: Echiniscidae) from the Republic of Ireland with comments on the 'weglarskae group' and other heterotardigrade taxa, including the controversial genus Bryochoerus Marcus, 1936. Journal of Animal Diversity 4, 1–52. http://dx.doi.org/10.52547/ JAD.2022.4.2.1
- de Zio Grimaldi, S., Gallo, M.D. and Lucia, M.R.M. 1992 Neoarctus primigenius n. g., n. sp., a new Stygarctidae of the Tyrrhenian Sea (Tardigrada, Arthrotardigrada). Bolletino di zoologia 59, 309–13. http://dx.doi. org/10.1080/11250009209386687
- Doyère, M. 1840 Mémoire sur les tardigrades. Annales des Sciences Naturelles 2, 269–361.
- Ehrenberg, C.G. 1853 Diagnoses novarum formarum. Bericht über die zur Bekanntmachung geeigneten Verhandlungen der Königlich Preussischen Akademie der Wissenschaften zu Berlin, 526–33.
- Fontoura P. and Morais, P. 2011 Assessment of traditional and geometric morphometrics for distinguishing cryptic species for the *Pseudechiniscus suillus* complex (Tardigrada, Echiniscidae). *Journal of Zoological Systematics and Evolutionary Research* **49**, 26–33. https:// doi.org/10.1111/j.1439-0469.2010.00594.x
- Fontoura, P., Pilato, G., Lisi, O. and Morais, P. 2009 Tardigrades from Portugal: four new records and description of two new species. *Zootaxa* 2030, 21–38. https://doi.org:10.11646/zootaxa.2030.1.2
- Fujimoto, S. and Hansen, J.G. 2019 Revision of Angursa (Arthrotardigrada: Styraconyxidae) with the description of a new species from Japan. European Journal of Taxonomy 510, 1–19. https://doi.org/10.5852/ ejt.2019.510
- Fujimoto, S. and Yamasaki, H. 2017 A new genus and species of Renaudarctidae (Heterotardigrada: Arthrotardigrada) from Ryukyu Archipelago, Japan. *Marine Biology Research* 13, 288–99. http://dx.doi.org/10.10 80/17451000.2016.1257809
- Gąsiorek, P. and Degma, P. 2018 Three Echiniscidae species (Tardigrada: Heterotardigrada) new to the Polish fauna, with the description of a new gonochoristic *Bryodelphax* Thulin, 1928. *Zootaxa* **4410**, 77–96. https://doi.org/10.11646/zootaxa.4410.1.4

- Gąsiorek, P., Morek, W., Stec, D. and Michalczyk, Ł. 2019a Untangling the *Echiniscus* Gordian knot: paraphyly of the "arctomys group" (Heterotardigrada: Echiniscidae). *Cladistics* 35, 633–53. https://doi.org/10.1111/ cla.12377
- Gąsiorek, P., Blagden, B. and Michalczyk, Ł. 2019b Towards a better understanding of echiniscid intraspecific variability: a redescription of *Nebularmis reticulatus* (Murray, 1905) (Heterotardigrada: Echiniscoidea). Zoologischer Anzeiger 283, 242–55. https:// doi.org/10.1016/j.jcz.2019.08.003
- Gąsiorek, P. and Michalczyk, Ł. 2020 Echiniscus siticulosus (Echiniscidae: spinulosus group), a new tardigrade from Western Australian scrub. New Zealand Journal of Zoology 47, 87–105. https://doi.org/10.1080/03014 223.2019.1603166
- Gąsiorek, P., Oczkowski, A., Blagden, B., Kristensen, R.M., Bartels, P.J., Nelson, D.R., Suzuki, A.C. and Michalczyk, Ł. 2021a New Asian and Nearctic Hypechiniscus species (Heterotardigrada: Echiniscidae) signalize a pseudocryptic horn of plenty. Zoological Journal of the Linnean Society 192, 794–852. https:// doi.org/10.1093/zoolinnean/zlaa110
- Gąsiorek, P., Stec, D., Morek, W., and Michalczyk, Ł. 2017 An integrative redescription of *Echiniscus te*studo (Doyère, 1840), the nominal taxon for the class Heterotardigrada (Ecdysozoa: Panarthropoda: Tardigrada). *Zoologischer Anzeiger* 270, 107–22. https://doi. org/10.1016/j.jcz.2017.09.006
- Gąsiorek, P., Stec, D., Zawierucha, K., Kristensen, R.M., and Michalczyk, Ł. 2018b Revision of *Testechiniscus* Kristensen, 1987 (Heterotardigrada: Echiniscidae) refutes the polar-temperate distribution of the genus. *Zootaxa* 4472, 261–97. https://doi.org/10.11646/ zootaxa.4472.2.3
- Gąsiorek, P., Suzuki, A.C., Kristensen, R.M., Lachowska-Cierlik, D. and Michalczyk Ł. 2018a Untangling the *Echiniscus* Gordian knot: *Stellariscus* gen. nov. (Heterotardigrada: Echiniscidae) from Far East Asia. *Invertebrate Systematics* **32**, 1234–47. https://doi. org/10.1071/IS18023
- Gąsiorek, P., Vončina, K., Zając, K. and Michalczyk, Ł. 2021b Phylogeography and morphological evolution of *Pseudechiniscus* (Heterotardigrada: Echiniscidae). *Scientific Reports* **11**, 7606. https://doi.org/10.1038/ s41598-021-84910-6
- Gąsiorek, P., Wilamowski, A., Vončina, K. and Michalczyk, Ł. 2022 Neotropical jewels in the moss: biodiversity, distribution and evolution of the genus *Barbaria* (Heterotardigrada: Echiniscidae). *Zoological Journal* of the Linnean Society **195**, 1037–66. https://doi. org/10.1093/zoolinnean/zlab087
- Greven, H., Kaya, M. and Baran, T. 2016 The presence of α-chitin in Tardigrada with comments on chitin in the Ecdysozoa. *Zoologischer Anzeiger* **264**, 11–16. http://dx.doi.org/10.1016/j.jcz.2016.06.003
- Grobys, D., Roszkowska, M., Gawlak, M., Kmita, H., Kepel, A., Kepel, M., Parnikoza, I., Bartylak, T. and Kaczmarek, Ł. 2020 High diversity in the *Pseudechiniscus suillus–facettalis* complex (Heterotardigrada: Echiniscidae) with remarks on the morphology of the genus *Pseudechiniscus. Zoological Journal of the Linnean Society* 188, 733–52. https://doi.org/10.1093/ zoolinnean/zlz171
- Guidetti, R., Rebecchi, L., Cesari, M. and McInnes, S. 2014 Mopsechiniscus franciscae, a new species of a rare

genus of Tardigrada from continental Antarctica. *Polar Biology* **37**, 1221–33. https://doi.org/10.1007/ s00300-014-1514-x

- Guil, N., Jørgensen, A., Giribet, G. and Kristensen, R.M. 2013 Congruence between molecular phylogeny and cuticular design in Echiniscoidea (Tardigrada, Heterotardigrada). Zoological Journal of the Linnean Society 169, 713–36. https://doi.org/10.1111/ zoj12090
- Hansen J.G., Kristensen R.M. and Jørgensen A. 2012 The armoured marine tardigrades (Arthrotardigrada, Tardigrada). Scientia Danica, Series B, Biologica 2, Det Kongelige Danske Videnskabernes Selskab.
- Jørgensen, A. 2000 Cladistic analysis of the Echiniscidae Thulin, 1928 (Tardigrada: Heterotardigrada: Echiniscoidea). Steenstrupia 25, 11–23.
- Jørgensen, A., Møbjerg. N. and Kristensen, R.M. 2011 Phylogeny and evolution of the Echiniscidae (Echiniscoidea, Tardigrada) – an investigation of the congruence between molecules and morphology. Journal of Zoological Systematics and Evolutionary Research **49**, 6–16. https://doi. org/10.1111/j.1439-0469.2010.00592.x
- Kayastha, P., Bartylak, T., Gawlak, M., and Kaczmarek, Ł. 2020 Integrative Description of *Pseudechiniscus lalitae* sp. nov. (Tardigrada: Heterotardigrada: Echiniscidae) from the Azores Archipelago (Portugal). *Annales Zoologici* **70**, 487–505. https://doi.org/10.3161/000345 41ANZ2020.70.4.002
- Kristensen, R.M. 1987 Generic revision of the Echiniscidae (Heterotardigrada) with a discussion of the origin of the family. In R. Bertolani (ed), Biology of Tardigrades. Selected Symposia and Monographs, 261– 335. Unione Zoologica Italiana.
- Kristensen, R.M. and Higgins, R.P. 1984 A new family of Arthrotardigrada (Tardigrada: Heterotardigrada) from the Atlantic coast of Florida, U.S.A. *Transactions* of the American Microscopical Society **103**, 295–311. https://doi.org/10.2307/3226191
- Kristensen, R.M. and Higgins, R.P. 1989 Marine Tardigrada from the Southeastern United States Coastal Waters I. *Paradoxipus orzeliscoides* n. gen., n. sp. (Arthrotardigrada: Halechiniscidae). *Transactions of the American Microscopical Society* **108**, 262–82. http:// dx.doi.org/10.2307/3226344
- Marcus, E. 1927 Zur Anatomie und Ökologie mariner Tardigraden. Zoologische Jahrbücher 53, 487–588.
- Marcus, E. 1936 Tardigrada. In R. Hesse, (ed), *Das Tierreich* 66, 1–340. Lieferung. Walter de Gruyter, Berlin.
- Maucci, W. 1986 Tardigrada, Fauna d'Italia 24. Bologna. Edizioni Calderini.
- Maucci, W. and Ramazzotti, G. 1981 Cornechiniscus gen. nov.: nuova posizione sistematica per i cosiddetti "Pseudechiniscus grouppo cornutus" con descrizione di una nuova specie (Tardigrada, Echiniscidae). Memorie dell'Istituto Italiano di Idrobiologia 39, 147–51.
- McInnes, S.J. 1994 Zoogeographic distribution of terrestrial/freshwater tardigrades from current literature. *Journal of Natural History* 28, 257–352. https://doi. org/10.1080/00222939400770131
- McInnes, S.J. Michalczyk, Ł. and Kaczmarek, Ł. 2017 Annotated zoogeography of non-marine Tardigrada. Part IV: Africa. *Zootaxa* 4284, 1–74. https://doi. org/10.11646/zootaxa.4284.1.1

- Mihelčič, F. 1951 Beitrag zur Systematik der Tardigraden. Archivio Zoologico Italiano **36**, 57–103.
- Miller, W.R., Horning, D.S. and Heatwole, H.F. 2001 Tardigrades of the Australian Antarctic: Macquarie Island, sub-Antarctica. *Zoologischer Anzeiger* 240, 475– 91. https://doi.org/10.1078/0044–5231–00057
- Murray, J. 1905 Tardigrada of the Scottish Lochs. Transactions of the Royal Society of Edinburgh 41, 677–98. https://doi.org/10.5962/bhl.title.12517
- Noda H. 1985 Description of a new subspecies of Angursa bicuspis Pollock (Heterotardigrada, Halechiniscidae) from Tanabe Bay, Japan. Publications of the Seto Marine Biological Laboratory 30, 269–76.
- Pérez-Pech, W.A., de Jesús-Navarrate, A., DeMilio, E., Anguas-Escalante, A. and Hansen, J.G. 2020 Marine Tardigrada from the Mexican Caribbean with the description of *Styraconyx robertoi* sp. nov. (Arthrotardigrada: Styraconyxidae). *Zootaxa* 4731, 492–508. http://dx.doi.org/10.11646/zootaxa.4731.4.3.
- Petersen, B. 1951 The tardigrade fauna of Greenland. *Meddelelser om Grønland* **150**, 1–94.
- Pilato, G., Binda, M.G. and Lisi, O. 2005 Remarks on some Echiniscidae (Heterotardigrada) from New Zealand with the description of two new species. *Zootaxa* 1027, 27–45. https://doi.org/10.11646/ zootaxa.1027.1.2
- Pilato, G., Fontoura, P., Lisi, O. and Beasley, C. 2008 New description of *Echiniscus scabrospinosus* Fontoura, 1982 and description of a new species of *Echiniscus* (Heterotardigrada) from China. *Zootaxa* 1856, 41–54. https://doi.org/10.11646/zootaxa.1931.1.6
- Pollock, L.W. 1979 Angursa bicuspis n. g., n. sp., a new marine arthrotardigrade from the western North Atlantic. Transactions of the American Microscopical Society 98, 558–62. https://doi.org/10.2307/3225907
- Ramazzotti, G. and Maucci, W. 1983 Il Phylum Tardigrada (III. edizione riveduta e aggiornata). Memorie dell' Istituto Italiano di Idrobiologia 41, 1–1016.
- Rebecchi, L., Altiero, T., Eibye-Jacobsen, J., Bertolani, R. and Kristensen, R.M. 2008 A new discovery of *Novechiniscus armadilloides* (Schuster, 1975) (Tardigrada, Echiniscidae) from Utah, USA with considerations on non-marine Heterotardigrada phylogeny and biogeography. *Organisms Diversity and Evolution* 8, 58–65. https://doi.org/10.1016/j.ode.2006.11.002
- Renaud-Mornant, J. 1974 Une nouvelle famille de Tardigrades marins abyssaux: les Coronarctidae fam nov (Heterotardigrada). Comptes Rendus Hebdomadaires des Séances de l'Académie des Sciences, Paris 278, 3087–90.
- Richters, F. 1904 Arktische Tardigraden. Fauna Arctica 3, 495–508.
- Richters, F. 1926 Tardigrada. In W. Kükenthal and T. Krumbach (eds) *Handbuch der Zoologie 3*. Berlin and Leipzig, Walter de Gruyter and Co.
- Roszkowska, M., Grobys, D., Bartylak, T., Gawlak, M., Kmita, H., Kepel, A., Kepel, M., Parnikoza, I. and Kaczmarek, Ł. 2020 Integrative description of five *Pseudechiniscus* species (Heterotardigrada: Echiniscidae: the *suillus-facettalis* complex). *Zootaxa* **4763**, 451–84. https://doi.org/10.11646/ zootaxa.4763.4.1
- Schultze, C.A.S. 1840 *Echiniscus Bellermanni; animal crustaceum.* Berolini. Apud G. Reimer.

- Schuster, R.O. 1975 A new species of *Parechiniscus* from Utah (Tardigrada: Echiniscidae). *Memorie dell'Istituto Italiano di Idrobiologia* **Suppl. 32**, 333–36.
- Scourfield, D.J. 1897 Contributions to the Non-Marine Fauna of Spitsbergen. Part I. Preliminary Notes, and Reports on the Rhizopoda, Tardigrada, Entomostraca, etc. Proceedings of the Zoological Society of London 65, 784–92. https://doi.org/10.1111/j.1096-3642.1897. tb03120.x
- Thulin, G. 1911 Beiträge zur Kenntnis der Tardigraden fauna Schwedens. Arkiv för Zoologi 7, 1–60. https:// doi.org/10.5962/bhl.part.1270.
- Thulin, G. 1928 Über die Phylogenie und das System der Tardigraden. *Hereditas* **11**, 207–66. https://doi. org/10.1111/j.1601-5223.1928.tb02488.x
- Tumanov, D.V. 2005 Isohypsibius panovi, a new species of Tardigrada from Ireland (Eutardigrada, Hypsibiidae). Zootaxa 812, 1–4. https://doi.org/10.11646/ zootaxa.812.1.1
- Tumanov, D.V. 2020 Analysis of non-morphometric morphological characters used in the taxonomy of the genus *Pseudechiniscus* (Tardigrada: Echiniscidae). *Zoological Journal of the Linnean Society* 188, 735–75. https://doi.org/10.1093/zoolinnean/zlz097